

Sex-specific inhibition and stimulation of worker-reproductive transition in a termite

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Abstract In social insects, the postembryonic development of individuals exhibits strong phenotypic plasticity in response to the environment, thus generating the caste system. Different from eusocial Hymenoptera, in which queens dominate reproduction and inhibit worker fertility, the primary reproductive caste in termites (kings and queens) can be replaced by neotenic reproductives derived from functionally sterile individuals. Feedback regulation of nestmate differentiation into reproductives has been suggested, but the sex specificity remains inconclusive. In the eastern subterranean termite, *Reticulitermes flavipes*, we tested the hypothesis that neotenic reproductives regulate worker-reproductive transition in a sex-specific manner. With this *R. flavipes* system, we demonstrate a sex-specific regulatory mechanism with both inhibitory and stimulatory functions. Neotenic inhibit workers of the same sex from differentiating into additional reproductives but stimulate workers of the opposite sex to undergo this transition. Furthermore, this process is not affected by the presence of soldiers. Our results highlight the reproductive plasticity of termites in response to social cues and provide insights into the regulation of reproductive division of labor in a hemimetabolous social insect.

Keywords Caste differentiation · Developmental plasticity · Ergatoid reproductive · *Reticulitermes flavipes*

Introduction

Developmental plasticity plays an important role in the reproductive division of labor in social insects (Page and Amdam 2007). Caste differentiation in eusocial colonies is usually dependent on social stimuli as well as other environmental cues (Hartfelder and Engels 1998; Korb and Hartfelder 2008). Although a fertilized egg is thought to be totipotent and able to develop into any caste, only a few individuals eventually become reproductives. For example, female honeybee larvae that are fed with royal jelly develop into queens, while others become workers (Kucharski et al. 2008). The presence of queens in social Hymenoptera also inhibits worker reproduction by directly suppressing their ovarian development or through policing behavior (Le Conte and Hefetz 2008).

As with most social insects, termites have caste systems resulting from developmental plasticity. In contrast with social Hymenoptera, hemimetabolous termites have both males and females for all castes. Termite colonies are typically founded by a pair of dispersing alates, which become the primary reproductives, i.e., kings and queens. In many “higher” termite genera (Termitidae) and most “lower” termite genera (all other termite families), workers and nymphs can differentiate into neotenic reproductives (ergatoids and nymphoids, respectively) and reproduce in the natal colony (Myles 1999; Roisin 2000; Roisin and Korb 2011). Neotenic reproduction is implicated to play a critical role in the early evolution of termite eusociality (Myles 1999). The fact that neotenic develop in response to orphaning (the absence of reproductives) has led to the prevailing hypothesis that fertile reproductives would inhibit sexual development (Long et al. 2003; Matsuura et al.

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2010; Moore 1974; Noirot 1990). A few studies, however, proposed the stimulatory effects of reproductive individuals on this process. For example, in *Mastotermes darwiniensis*, neotenic reproductives were produced in the presence rather than the absence of neotenics, and female neotenics exhibited stronger stimulatory activities on workers of both sexes than males (Watson et al. 1975). In *Kaloterms flavicollis*, the production of female neotenics was promoted by the presence of a single male neotenic, while the stimulatory effect was not observed from female neotenics (Lüscher 1964). Although kings and queens can both be replaced by neotenics, the sex specificity for either inhibition or potential stimulation is not conclusive in termites.

Reticulitermes, one of the most widely distributed termite genera in the world with substantial economic and ecological importance (Su et al. 2001), is an ideal system to study developmental plasticity. *Reticulitermes* workers have three morphologically, behaviorally and functionally distinct developmental trajectories. They can undergo *status quo* molts and remain as workers, differentiate into pre-soldiers followed by an additional molt into soldiers, or develop into neotenic reproductives (i.e., ergatoids) (Lainé and Wright 2003; Zhou et al. 2006). Our preliminary study in the eastern subterranean termite *Reticulitermes flavipes* indicated that worker-reproductive transition was a lengthy process under orphaning condition (30–90 days). If one of the reproductives (e.g., queen) is lost, a stimulatory function from the remaining reproductive (e.g., king) that promotes the formation of neotenic reproductives of the missing sex (e.g., female ergatoid) would be beneficial to the colony. We hypothesized that worker-reproductive transition is regulated in a sex-specific manner in *R. flavipes*. Specifically, reproductive individuals inhibit same-sex workers but stimulate opposite-sex workers to differentiate into ergatoids. To test this, we evaluated ergatoid formation in response to the presence or absence of male or female reproductives. As a soldier caste is present in all termite species (Tian and Zhou 2014), and previous studies suggest that soldiers potentially promote differentiation of reproductives (Watanabe et al. 2014), we also examined the effect of soldiers on ergatoid formation.

Methods

Study system

Colonies of *R. flavipes* were collected from the Arboretum (Lexington, Kentucky, USA) and the Red River Gorge area, Daniel Boone National Forest (Slade, Kentucky, USA). Colonies consisted of workers, soldiers, and nymphs at the time of collection. Freshly collected termites were kept in Petri dishes and fed on moistened unbleached paper towel for 1 to 2 weeks before being further used. Neotenic-headed colonies were obtained by transferring field collected termites to closed plastic boxes

(45.7 × 30.6 × 15.2 cm) filled with moistened wood mulch and pinewood blocks and maintained for 6 months, when eggs and larvae appeared indicating the presence of fertile neotenic reproductives. Primary-headed colonies were established by pairing female and male sibling alates collected in Lexington, Kentucky, and they had been maintained in laboratory for 5 years by the time experiments started. All stock colonies and experimental termites were maintained at 27 ± 1 °C in complete darkness.

Bioassay to test sex-specific regulation

Fertile ergatoids of different sexes were used to test their influences on worker-reproductive transition. These ergatoids were obtained from an orphaning assay, in which groups of 100 workers were kept in Petri dishes (6.0 cm in diameter, 1.5 cm in height) lined with moistened unbleached paper towel for 60 days. Ergatoids that were actively reproducing (i.e., with eggs present in dishes) were used in the subsequent assay.

The same set-up was used to test how workers differentiate in response to ergatoids (Fig. 1). The bioassay was conducted according to Matsuura et al. (2010) with modification. Specifically, groups of 100 workers were kept with (1) no reproductives (F–M–); (2) one female ergatoid (F+ M–); (3) one male ergatoid (F–M+); or (4) one pair of ergatoids (F+ M+). The ergatoids and workers of each group were originally from the same colony. Fourteen replications using three colonies were conducted, with one colony originally primary-headed (four replications) and two colonies originally neotenic-headed (five replications each). The differentiation of workers was recorded for 60 consecutive days, and newly formed ergatoids were removed every day to prevent their potential inhibition on differentiation of additional ergatoids, and aggression against ergatoids formed later. Ergatoids were recognized by slightly heavier cuticle pigmentation, elongated abdomen, and wider thorax than workers (Fig. 2). Female ergatoids were distinguished from males by their enlarged 7th abdominal sternite. We also removed eggs, newly formed pre-soldiers, and any inter-caste (individuals with the degree of mandible development intermediate between workers and pre-soldiers) every day to prevent their potential influence on worker development.

Bioassays to test soldier effect

Two orphaning assays were conducted to test the influence of soldier caste on ergatoid differentiation. The first assay (“single-soldier orphaning assay”) simulated the natural orphaning condition where freshly collected workers were separated into groups of 100 individuals, and each group was placed with one soldier (soldier+) or no soldier (soldier–). Termites were maintained in Petri dishes (6.0 cm in diameter, 1.5 cm in height) lined with moistened unbleached paper towel as food source. Termites were allowed to undergo changes in caste differentiation in the dishes without disturbance, and caste composition and mortality

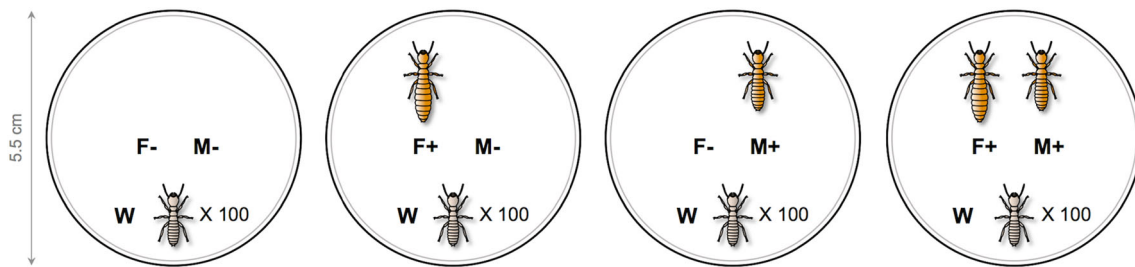


Fig. 1 Experimental set-up. Each group of 100 workers was placed in a Petri dish lined with moistened unbleached paper. Workers were kept with no reproductives ($F^- M^-$), one female ergatoid ($F^+ M^-$), one male ergatoid ($F^- M^+$), or one pair of ergatoids ($F^+ M^+$). Newly formed ergatoids, pre-soldiers, and eggs laid by reproductives were

removed every day for 60 consecutive days. F female ergatoid reproductive, M male ergatoid reproductive, W worker. Ergatoid reproductives used in the assay were actively reproducing, and female ergatoids were physogastric

of each group were documented at the end of 60 days. A total of 42 and 45 replicates from four colonies were conducted for soldier+ and soldier- treatment, respectively.

We further performed the second assay (“multiple-soldier orphaning assay”), which was conducted with an increased soldier stimulus and observed daily for 60 consecutive days. Groups of 100 workers were isolated from neotenic-headed colonies, and they were companioned with either four soldiers (soldier++) or no soldier (soldier-). Newly differentiated individuals (ergatoids and pre-soldiers) were removed to prevent their potential influence on worker development, and an equal number of workers to the removed individuals were added to the group to keep group size consistent. Termites were maintained in Petri dishes (3.5 cm in diameter, 1.5 cm in height) provided with moistened unbleached paper towel. A total of ten replicates from two colonies were conducted for both soldier++ and soldier- treatments.

Statistical analyses

Data were analyzed using Statistix 10 (Analytical Software, Tallahassee, FL, USA). In the assay that tested sex-specific

regulation, Wilcoxon rank-sum test was performed on the cumulative numbers of ergatoids between each treatment and the control on every assay day. In both single- and multiple-soldier orphaning assays, unpaired t test was performed on numbers of differentiated individuals and mortality. To obtain values that fit the assumptions of parametric test, data were transformed through square root ($x + 0.5$) on the combined numbers of pre-soldiers and soldiers in the single-soldier orphaning assay, and numbers of female and male ergatoids in the multiple-soldier orphaning assay. Because the pattern of regulation was consistent in all colonies, results were pooled for statistical analyses.

Results

Ergatoid formation in response to reproductives of different sexes

Under orphaning condition ($F^- M^-$), 4.1 ± 1.3 and $2.5 \pm 0.7\%$ of workers differentiated into female and male ergatoids, respectively, in 60 days (mean \pm SEM, Fig. 3). The presence of a single female ($F^+ M^-$) or a pair of ergatoids ($F^+ M^+$) significantly

Fig. 2 Photographs of a worker, young ergatoids, and mature ergatoids. The young ergatoids were about 10 days postdifferentiation. The mature ergatoids were 6 months postdifferentiation and actively reproducing

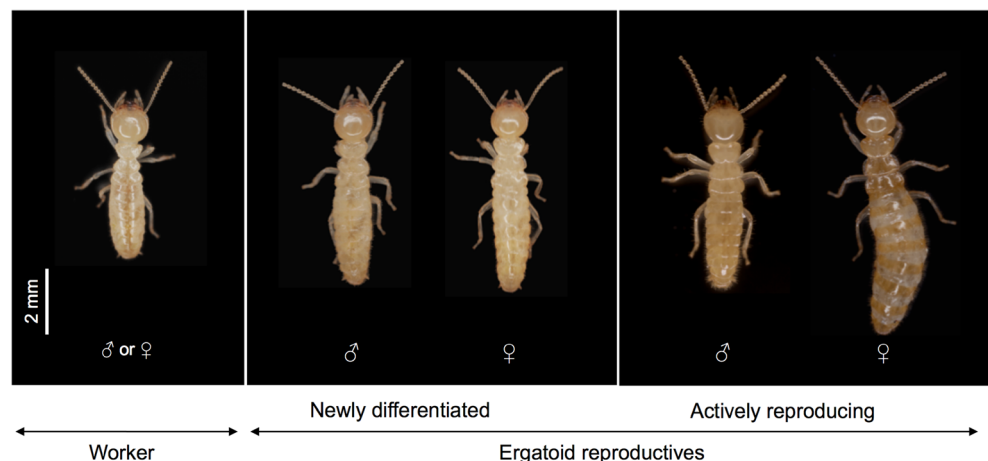


Fig. 3 Ergatoid formation in response to fertile ergatoids of different sexes. Cumulative percentage of newly differentiated female (a) and male (b) ergatoids is shown (mean \pm SEM). Stimulation (*forward arrows*) and inhibition (*reverse arrows*) refer to significantly more and fewer ergatoids formed, respectively, relative to control ($F^- M^-$), and *dashed line next to the tip of arrow* indicates the day when the significant difference started (Wilcoxon rank-sum test, one-sided, $P < 0.01$; $n = 14$ for all treatments). *Symbols and arrows of the same color correspond to the same treatment*

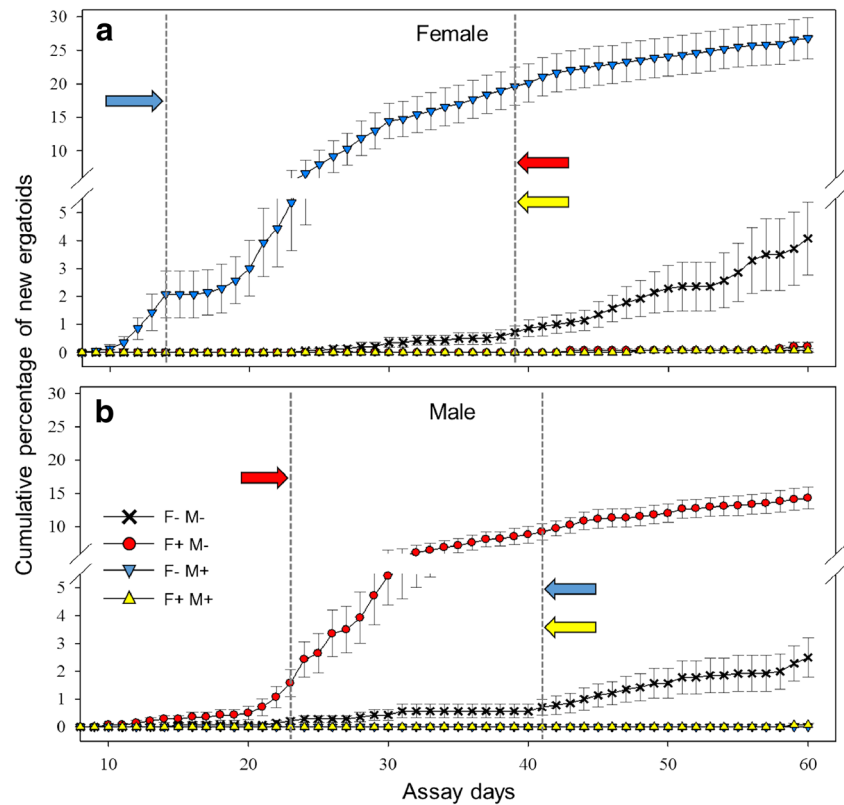
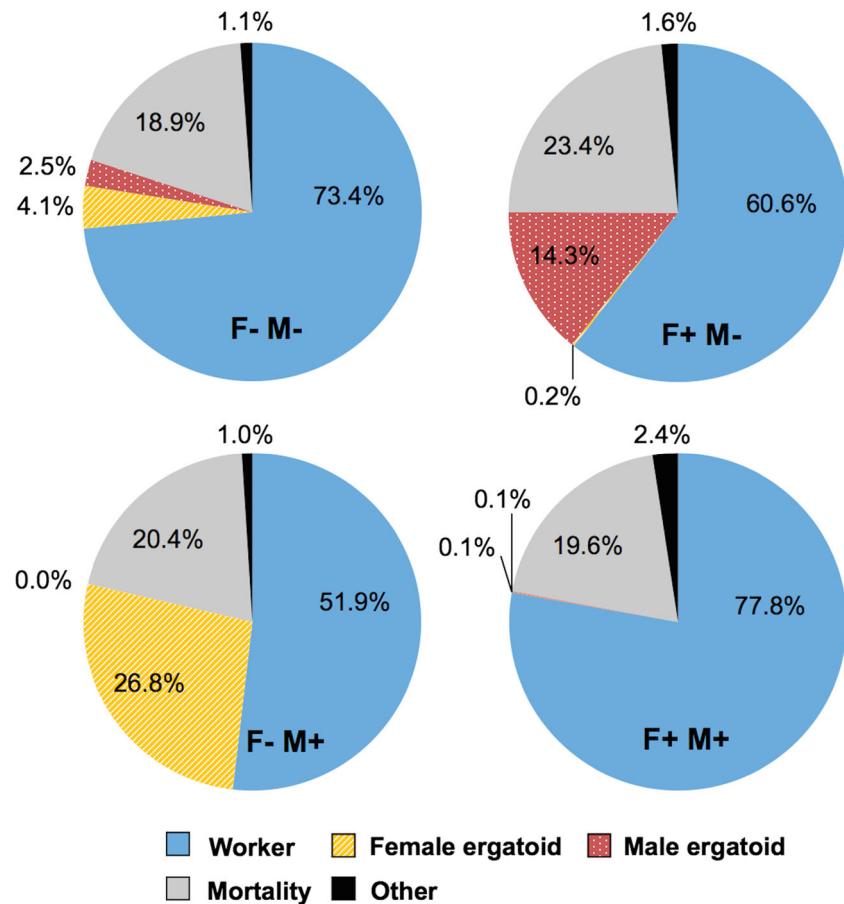


Fig. 4 The developmental endpoint of workers in 60 days. *Other* includes pre-soldiers and inter-castes, and the latter refers to individuals with the degree of mandible development between workers and pre-soldiers after molting. There was no significant effect of treatment on mortality (ANOVA, $F_{3, 52} = 1.5$, $P > 0.05$; $n = 14$ pooled from three colonies)



inhibited the formation of additional female ergatoids ($0.2 \pm 0.2\%$ and $0.1 \pm 0.1\%$, respectively); however, the presence of a single male ergatoid (F– M+) significantly stimulated the formation of female ergatoid ($26.8 \pm 3.0\%$) (Fig. 3a; $P < 0.01$, Wilcoxon rank-sum test, one-sided, $n = 14$). Similarly, significantly fewer male ergatoids differentiated in the presence of a single male (F– M+) or a pair of ergatoids (F+ M+) (0 and $0.1 \pm 0.1\%$, respectively), while significantly more of them formed when a single female ergatoid was present (F+ M–) ($14.3 \pm 1.6\%$) (Fig. 3b; $P < 0.01$, Wilcoxon rank-sum test, one-sided, $n = 14$).

Within 60 days, female and male ergatoids differentiated in 10 and 9 replicates, respectively, out of 14 total replicates under the orphaning condition (F– M–). The formation of ergatoids in all 14 replicates was stimulated by a single ergatoid of the opposite sex. Under this stimulation, developmental time for the first ergatoid was significantly reduced (female, 19 ± 3.3 days, $n = 14$ in F– M+, compared with 38 ± 3.7 days, $n = 10$ in F– M–; male, 21 ± 1.8 days, $n = 14$ in F+ M–, compared with 35 ± 5.4 days, $n = 9$ in F– M–; Wilcoxon rank-sum test, one-sided, $P < 0.01$). There was no significant difference on mortality within 60-day assay period among treatments (Fig. 4; ANOVA, $F_{3, 52} = 1.5$, $P > 0.05$; $n = 14$).

Ergatoid formation in response to soldier caste

In both assays, ergatoids were differentiated from workers in response to orphaning condition, and there were no significant effects of soldiers on ergatoid formation (Fig. 5). In the single-soldier orphaning assay, soldier caste did not significantly influence the number of female or male ergatoids in 60 days (Fig. 5a; female: $t_{85} = 1.64$, $P > 0.05$; male: $t_{85} = 1.64$, $P > 0.05$; unpaired t test, two-sided, $n = 42$ for soldier+, $n = 45$ for soldier–). At the end of the assay, mortality between soldier+ and soldier– groups were not significantly different (Fig. 5b; $t_{42} = 0.67$, $P > 0.05$; unpaired t test, two-sided, $n = 22$ randomly selected from both soldier+ and soldier– groups). The presence of one soldier significantly suppressed the differentiation of pre-soldiers and soldiers, and the increased total number of pre-soldiers and soldiers are 0.43 ± 0.10 in soldier+ groups and 0.91 ± 0.14 in soldier– groups ($t_{85} = 2.71$, $P < 0.01$; unpaired t test, two-sided; data were transformed (square root ($x + 0.5$))); $n = 42$ for soldier+, $n = 45$ for soldier–; Fig. S1). Similarly, in the multiple-soldier orphaning assay, the soldier caste did not significantly influence the accumulative number of female or male ergatoids in 60 days (Fig. 5c; female: $t_{18} = 0.57$, $P > 0.05$; male: $t_{18} = 0.44$, $P > 0.05$; unpaired t test, two-sided; data were transformed (square root ($x + 0.5$))); $n = 10$ for both soldier++ and soldier– groups). Mortality was

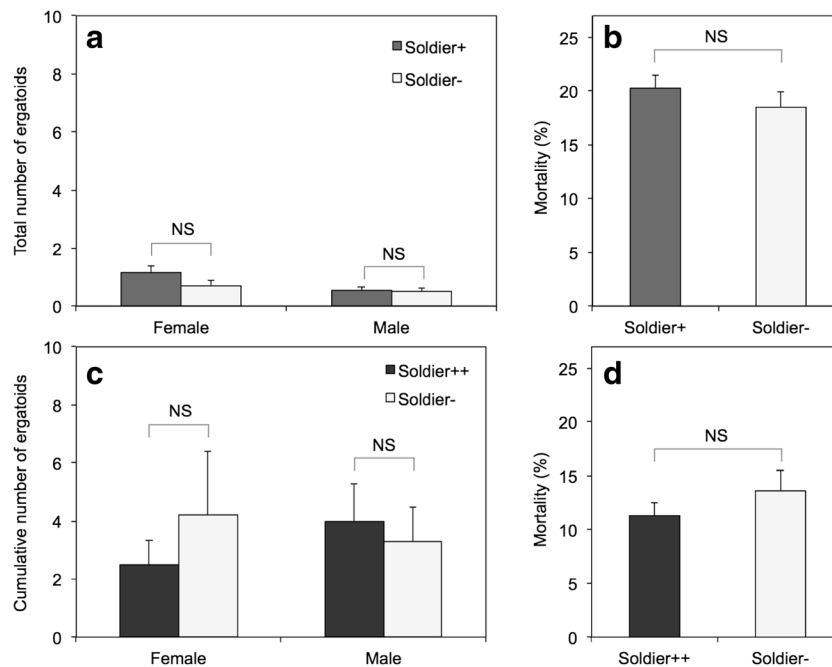


Fig. 5 Influence of soldiers on ergatoid formation. In a single-soldier orphaning assay, total numbers of female and male ergatoids presented (mean \pm SEM) (a) and mortality (%; mean \pm SEM) (b) between soldier present and absent groups in 60 days are shown. In a multiple-soldier orphaning assay, cumulative numbers of female and male ergatoids differentiated (mean \pm SEM) (c) and mortality (%; mean \pm SEM) (d) between soldier present and absent groups in 60 days are shown. In the single-soldier assay, newly differentiated ergatoids were left in groups; soldier+ each group consisted of 100 workers and one soldier at the start

of assay, soldier– each group consisted of 100 workers only, NS no significant difference (unpaired t test, two-sided, $P > 0.05$; $n = 42$ for soldier+, $n = 45$ for soldier–). In the multiple-soldier assay, newly differentiated ergatoids were removed and replaced with workers every day; soldier++ each group consisted of 100 workers and four soldiers at the start of assay, soldier– each group consisted of 100 workers only, NS no significant difference (unpaired t test, two-sided, $P > 0.05$; $n = 10$ for both soldier++ and soldier–)

not significantly influenced by the presence of four soldiers (Fig. 5d; $t_{18} = 1.02$, $P > 0.05$; unpaired t test, two-sided, $n = 10$ for both soldier++ and soldier− groups).

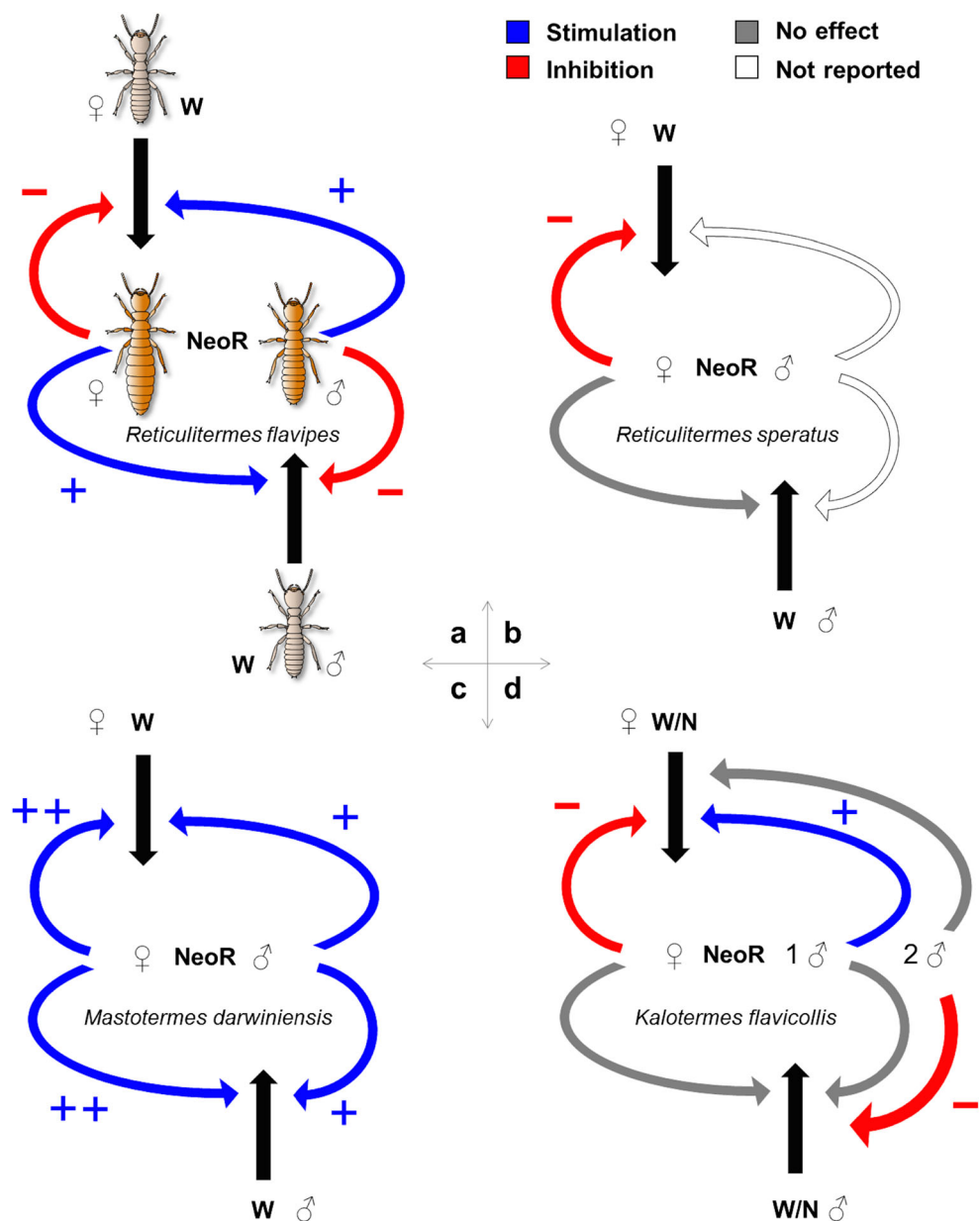
Discussion

These results support our hypothesis that regulation of worker-reproductive transition by fertile reproductives is sex-specific. More importantly, our empirical evidence demonstrated that the dual regulation (inhibition and stimulation) is employed by both sexes. Ergatoid differentiation occurs after more than 30 days in response to orphaning but can be significantly accelerated by the presence of a potential mate.

Such stimulation by the opposite sex benefits the colony by enabling it to resume reproduction soon after the loss of a queen or a king. Inhibition of development by the same sex, on the other hand, prevents unnecessary investments in reproduction, which, in turn, would be a loss in the labor force. This sex-specific regulation suggests that the development of reproductives is strictly dependent on the reproductive needs of the colony.

Neotenic reproduction is common in termites; however, regulation of neotenic differentiation varies among species (Grassé and Noirot 1960; Lüscher 1964; Matsuura et al. 2010; Miyaguni et al. 2013; Watson et al. 1975). In *Reticulitermes speratus*, female reproductives inhibit the differentiation of female neotenics but do not influence the

Fig. 6 Comparison of feedback regulation of neotenic differentiation in four termites. **a** Sex-specific inhibition and stimulation are demonstrated for both females and males in *Reticulitermes flavipes* (this study). **b** In *Reticulitermes speratus*, female neotenics inhibit differentiation of females but does not influence males; the effects of male neotenics were not reported. **c** In *Mastotermes darwiniensis*, both female and male neotenics stimulate neotenic differentiation but not in a sex-specific manner; females exhibit stronger stimulation. **d** In *Kaloterme flavicollis*, female neotenics inhibit differentiation of females, but the effect of males depends on the number. One male neotenic shows opposite-sex stimulation, while two males exhibit same-sex inhibition. W worker, N nymph, NeoR neotenic reproductive



formation of male neotenic (Matsuura et al. 2010). Compared with *R. flavipes*, female ergatoid formation is faster in *R. speratus* in response to orphaning, and the formation of nymphoids are faster than ergatoids in *R. speratus* (Matsuura et al. 2010; Miyata et al. 2004). Orphaning assays have also been conducted in other congeneric species including *R. grassei* (Pichon et al. 2007) and *R. urbis* (Ghesini and Marini 2009), which confirmed the inhibitory effect of reproductive pairs, but the regulation by each sex remains unclear. Head butting by workers is a behavioral indicator of reproductive disinhibition (Korb et al. 2009). In the damp wood termite *Zootermopsis nevadensis*, female and male workers equally increased head-butting behavior regardless of the sex of the reproductives removed, indicating the lack of sex specificity in this species (Penick et al. 2013). Stimulation by neotenic reproductives has been suggested previously in some primitive termite species. In *K. flavicollis*, the formation of female neotenic was stimulated by the extracts of male reproductives (Lüscher 1964). In *M. darwiniensis*, the formation of neotenic was promoted in the presence rather than the absence of other neotenic. Although sex specificity was not confirmed, female neotenic exhibited stronger stimulatory effects than males and the pair (Watson and Abbey 1985; Watson et al. 1975). In comparison, *R. flavipes* neotenic exhibit sex-specific inhibition and stimulation in both sexes (Fig. 6a). Such a regulatory pattern consists of all possible directions of social regulation, therefore presents a model for understanding the pheromonal and developmental mechanisms underlying neotenic reproduction. Given that previous studies on the differentiation of neotenic reproductives were incomplete or inconclusive (Fig. 6b–d), this study also provides an opportunity for us to re-examine the sex-specificity hypothesis across termite taxa.

Foraging populations of *R. flavipes* contain about 2% or fewer soldiers (Haverty and Howard 1981; Howard and Haverty 1980), while higher proportions (close to 4%) were observed in nest areas where neotenic are present (Howard and Haverty 1980). Soldiers were considered to induce the differentiation of reproductives (Tian and Zhou 2014; Watanabe et al. 2014); however, our results indicated that soldier caste does not play a significant role in regulating ergatoid formation in *R. flavipes*. It is worth noting that if ergatoids were not removed from the groups (single-soldier orphaning assay), the total number of ergatoids was lower than the cumulative number of ergatoids if they were constantly removed (multiple-soldier orphaning assay) within the same period of time. Although the two assays were conducted separately, this result could be explained by newly formed ergatoids suppressing formation of additional ergatoids through pheromones or policing behavior.

Our study represents the first step in understanding sex-specific worker-reproductive differentiation in response to social cues. The results of this study add a new dimension to the

prevailing view that reproductives inhibit worker-reproductive transition in termites (Noirot 1990). Much remains to be investigated about the regulatory mechanisms of caste differentiation, including the identification of inhibitory and stimulatory pheromones from reproductives. The active substances or blends must be sex-specific. The search of reproductive pheromones in termites should include volatile compounds (Matsuura et al. 2010), cuticular hydrocarbons (Liebig et al. 2009) as often observed in Hymenoptera (Van Oystaeyen et al. 2014), and proteinaceous secretions (Hanus et al. 2010). The sex specificity and the dual effect of reproductive cues reflect unique adaptation and regulation of caste differentiation in hemimetabolous termites.

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Author contributions Q.S., K.F.H., and X.Z. designed the experiments; Q.S. and J.D.H. conducted the experiments; Q.S. and K.F.H. analyzed the data; Q.S. wrote the manuscript; and K.F.H. and X.Z. revised the manuscript. All authors approved the final manuscript.

Compliance with ethical standards

Data availability All data generated or analyzed during this study are included in this published article and its supplementary information files.

References

- Ghesini S, Marini M (2009) Caste differentiation and growth of laboratory colonies of *Reticulitermes urbis* (Isoptera, Rhinotermitidae). *Insect Soc* 56:309–318
- Grassé PP, Noirot C (1960) Role respectif des males et des femelles dans la formation des sexués néoténiques chez *Calotermes flavicollis*. *Insect Soc* 7:109–123
- Hanus R, Vrkošlav V, Hrdý I, Cvačka J, Šobotník J (2010) Beyond cuticular hydrocarbons: evidence of proteinaceous secretion specific to termite kings and queens. *Proc R Soc B* 277:995–1002
- Hartfelder K, Engels W (1998) Social insect polymorphism: hormonal regulation of plasticity in development and reproduction in the honeybee. In: Pedersen RA, Schatten GP (eds) Current topics in developmental biology. Academic Press, San Diego, pp 45–78
- Haverty M, Howard R (1981) Production of soldiers and maintenance of soldier proportions by laboratory experimental groups of *Reticulitermes flavipes* (Kollar) and *Reticulitermes virginicus* (Banks) (Isoptera: Rhinotermitidae). *Insect Soc* 28:32–39

- Howard RW, Haverty MI (1980) Reproductives in mature colonies of *Reticulitermes flavipes*: abundance, sex-ratio, and association with soldiers. *Environ Entomol* 9:458–460
- Korb J, Hartfelder K (2008) Life history and development—a framework for understanding developmental plasticity in lower termites. *Biol Rev* 83:295–313
- Korb J, Weil T, Hoffmann K, Foster KR, Rehli M (2009) A gene necessary for reproductive suppression in termites. *Science* 324:758–758
- Kucharski R, Maleszka J, Foret S, Maleszka R (2008) Nutritional control of reproductive status in honeybees via DNA methylation. *Science* 319:1827–1830
- Lainé LV, Wright DJ (2003) The life cycle of *Reticulitermes* spp. (Isoptera: Rhinotermitidae): what do we know? *Bull Entomol Res* 93:267–278
- Le Conte Y, Hefetz A (2008) Primer pheromones in social Hymenoptera. *Annu Rev Entomol* 53:523–542
- Liebig J, Eliyahu D, Brent C (2009) Cuticular hydrocarbon profiles indicate reproductive status in the termite *Zootermopsis nevadensis*. *Behav Ecol Sociobiol* 63:1799–1807
- Long CE, Thorne BL, Breisch NL (2003) Termite colony ontogeny: a long-term assessment of reproductive lifespan, caste ratios and colony size in *Reticulitermes flavipes* (Isoptera: Rhinotermitidae). *Bull Entomol Res* 93:439–445
- Lüscher M (1964) Die spezifische Wirkung männlicher und weiblicher Ersatzgeschlechtstiere auf die Entstehung von Ersatzgeschlechtstieren bei der Termiten *Kaloterme flavicollis* (Fabr.) *Insect Soc* 11:79–90
- Matsuura K, Himuro C, Yokoi T, Yamamoto Y, Vargo EL, Keller L (2010) Identification of a pheromone regulating caste differentiation in termites. *Proc Natl Acad Sci U S A* 107:12963–12968
- Miyaguni Y, Sugio K, Tsuji K (2013) The unusual neotenic system of the Asian dry wood termite, *Neotermes koshunensis* (Isoptera: Kalotermitidae). *Sociobiology* 60:65–68
- Miyata H, Furuichi H, Kitade O (2004) Patterns of neotenic differentiation in a subterranean termite, *Reticulitermes speratus* (Isoptera: Rhinotermitidae). *Entomol Sci* 7:309–314
- Moore B (1974) Pheromones in the termite societies. In: Birch M (ed) *Pheromones*. North-Holland Publishing, Amsterdam, pp 250–266
- Myles TG (1999) Review of secondary reproduction in termites (Insecta: Isoptera) with comments on its role in termite ecology and social evolution. *Sociobiology* 33:1–43
- Noirot C (1990) Sexual castes and reproductive strategies in termites. In: Engels W (ed) *Social insects*. Springer, Berlin, pp 5–35
- Page RE, Amdam GV (2007) The making of a social insect: developmental architectures of social design. *BioEssays* 29:334–343
- Penick CA, Trobaugh B, Brent CS, Liebig J (2013) Head-butting as an early indicator of reproductive disinhibition in the termite *Zootermopsis nevadensis*. *J Insect Behav* 26:23–34
- Pichon A, Kutnik M, Leniaud L, Darrouzet E, Chaline N, Dupont S, Bagnères A (2007) Development of experimentally orphaned termite worker colonies of two *Reticulitermes* species (Isoptera: Rhinotermitidae). *Sociobiology* 50:1015–1034
- Roisin Y (2000) Diversity and evolution of caste patterns. In: Abe T, Bignell DE, Higashi M (eds) *Termites: evolution, sociality, symbioses, ecology*. Springer, Netherlands, pp 95–119
- Roisin Y, Korb J (2011) Social organisation and the status of workers in termites. In: Bignell DE, Roisin Y, Lo N (eds) *Biology of termites: a modern synthesis*. Springer, Netherlands, pp 133–164
- Su N-Y, Scheffrahn RH, Cabrera BJ (2001) Native subterranean termites: *Reticulitermes flavipes* (Kollar), *Reticulitermes virginicus* (Banks), *Reticulitermes hageni* Banks (Insecta: Isoptera: Rhinotermitidae). University of Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, EDIS, Gainesville
- Tian L, Zhou X (2014) The soldiers in societies: defense, regulation, and evolution. *Int J Biol Sci* 10:296–308
- Van Oystaeyen A, Oliveira RC, Holman L et al (2014) Conserved class of queen pheromones stops social insect workers from reproducing. *Science* 343:287–290
- Watanabe D, Gotoh H, Miura T, Maekawa K (2014) Social interactions affecting caste development through physiological actions in termites. *Front Physiol* 5:127
- Watson J, Abbey HM (1985) Development of neotenic in *Mastotermes darwiniensis* Froggatt: an alternative strategy. In: Watson JAL, Okot-Kotber BM, Noirot C (eds) *Caste differentiation in social insects*. Pergamon Press, Oxford, pp 107–124
- Watson JAL, Metcalf EC, Sewell JJ (1975) Preliminary studies on the control of neotenic formation in *Mastotermes darwiniensis* Froggatt (Isoptera). *Insect Soc* 22:415–426
- Zhou X, Oi FM, Scharf ME (2006) Social exploitation of hexamerin: RNAi reveals a major caste-regulatory factor in termites. *Proc Natl Acad Sci U S A* 103:4499–4504